

(19) World Intellectual Property Organization  
International Bureau



(43) International Publication Date  
13 June 2002 (13.06.2002)

PCT

(10) International Publication Number  
**WO 02/46451 A2**

(51) International Patent Classification<sup>7</sup>: **C12Q 1/00**

(21) International Application Number: **PCT/US01/50890**

(22) International Filing Date: 25 October 2001 (25.10.2001)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:  
09/697,225 26 October 2000 (26.10.2000) US

(71) Applicant (for all designated States except US): **PARADIGM GENETICS, INC.** [US/US]; 104 Alexander Drive, Bldg. 2, P.O. Box 14528, Research Triangle Park, NC 27709-4528 (US).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **KLOTI, Andreas** [CH/CH]; Buechlenweg 12, CH-8805 Richterswil (CH). **WOESSNER, Jeffrey** [US/US]; 2728 Canter Drive, Hillsborough, NC 27278 (US). **ZAYED, Adel** [EG/US]; 1021 Fenwick Parkway, Durham, NC 27713 (US). **BOYES, Douglas** [US/US]; 361 Vickers Road, Chapel Hill, NC 27514 (US). **DAVIS, Keith** [US/US]; 4119 Cobblestone Place, Durham, NC 27707 (US). **HAMILTON, Carol** [US/US]; 2006 Weehawken Place, Apex, NC 27502 (US). **ASCENZI, Robert** [US/US]; 741 Landing Lane, Cary, NC 27511 (US). **HOFFMAN, Neil** [US/US]; 115 Silo Drive, Chapel Hill, NC 27514 (US).

(74) Agents: **HOFMEYER, Timothy, G. et al.**; Paradigm Genetics, Inc., 104 Alexander Drive, Building 2, P.O. Box 14528, Research Triangle Park, NC 27709-4528 (US).

(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

**Published:**

- without international search report and to be republished upon receipt of that report
- with sequence listing part of description published separately in electronic form and available upon request from the International Bureau

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: METHODS FOR THE IDENTIFICATION OF INHIBITORS OF CYSTEINE SYNTHASE EXPRESSION OR ACTIVITY IN PLANTS

(57) Abstract: The present inventors have discovered that cysteine synthase is essential for plant growth. Specially, the inhibition of cysteine synthase gene expression in plant seedlings results in severe developmental abnormalities. Thus, cysteine synthase can be used as a target for the identification of herbicides. Accordingly, the present invention provides methods for the identification of compounds that inhibit cysteine synthase expression or activity, comprising: contacting a compound with a cysteine synthase and detecting the presence and/or absence of binding between said compound and said a cysteine synthase, or detecting a decrease in cysteine synthase expression or activity. The methods of the invention are useful for the identification of herbicides.

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## METHODS FOR THE IDENTIFICATION OF INHIBITORS OF CYSTEINE SYNTHASE EXPRESSION OR ACTIVITY IN PLANTS

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### FIELD OF THE INVENTION

The invention relates generally to plant molecular biology. In particular, the invention relates to methods for the identification of herbicides.

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### BACKGROUND OF THE INVENTION

Cysteine synthase (EC 4.2.99.8) is an enzyme involved in the final step of cysteine biosynthesis and catalyzes the formation of the amino acid L-cysteine from  
15 O-acetyl-L-serine (OAS) and inorganic sulfide. Cysteine synthase genes have been characterized from a number of plants, and in most cases consist of a small multigene family. At least ten different cysteine synthase genes have been identified in *Arabidopsis thaliana*. The *Arabidopsis* cysteine synthase gene, *AtcysC1*, encodes a  
mitochondrial protein having both cysteine synthase activity (EC 4.2.99.8) and  $\beta$ -  
20 cyanoalanine synthase activity (EC 4.4.1.9). Yamaguchi *et al.* (2000) *Plant Cell Physiol* 41:465-476.

To date, there has been no report or suggestion that cysteine synthase activity is essential for plant growth and development. Rather, there have been several reports that inhibition of cysteine synthase expression had no effect on plant growth. For  
25 example, constitutive antisense RNA expression of a spinach cytoplasmic cysteine synthase (CSase A) cDNA in tobacco plants results in decreased cysteine synthase activity, but did not damage transgenic plants. Saito (1994) *Plant Physiol* 106:887-

895. Furthermore, constitutive antisense RNA expression of a wheat cytoplasmic cysteine synthase (*cysI*) cDNA in tobacco plants had no effect on cysteine synthase activity or on the phenotype of the transgenic plant. Thus, the prior art has not suggested that cysteine synthase is essential for plant growth and development.

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## SUMMARY OF THE INVENTION

Surprisingly, the present inventors have discovered that antisense expression of cysteine synthase cDNA in *Arabidopsis* results in the production of white seeds that fail to germinate. Thus, the present inventors have discovered that cysteine synthase is essential for normal seed development and germination, and can be used as a target for the identification of herbicides. Accordingly, the present invention provides methods for the identification of compounds that inhibit cysteine synthase expression or activity, comprising: contacting a compound with a cysteine synthase and detecting the presence and/or absence of binding between said compound and said cysteine synthase, or detecting a decrease in cysteine synthase expression or activity. The methods of the invention are useful for the identification of herbicides.

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## DETAILED DESCRIPTION OF THE INVENTION

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### Definitions

The term "binding" refers to a noncovalent interaction that holds two molecules together. For example, two such molecules could be an enzyme and an inhibitor of that enzyme. Noncovalent interactions include hydrogen bonding, ionic interactions among charged groups, van der Waals interactions and hydrophobic interactions among nonpolar groups. One or more of these interactions can mediate the binding of two molecules to each other.

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Cysteine is described as compound No. 2850 in The Merck Index, 12<sup>th</sup> Ed., Merck Research Laboratories, Whitehouse Station, NJ, 1996. The term cysteine is synonymous with L-cysteine, Cys,  $\beta$ -mercaptoalanine, (R)-2-amino-3-mercaptopropanoic acid, 2-amino-3-mercaptopropionic acid,  $\alpha$ -amino- $\beta$ -thiolpropionic acid, half-cysteine and thioserine.

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As used herein, the term "cysteine synthase" (EC 4.2.99.8) is synonymous with "O-acetyl-L-serine (thiol) lyase", "O-acetylserine lyase", "O-acetyl-L-serine lyase", "O-acetyl-L-serine sulphydrylase" and "OAS sulphydrylase". For the purposes of the invention, cysteine synthase refers to any enzyme that catalyzes the formation of amino acid L-cysteine and acetate from O-acetylserine and H<sub>2</sub>S. Thus, the Arabidopsis enzyme referred to as cysteine synthase C (AtCS-C, Csase C) and encoded by *AtcysC1*, which has both beta-cyanoalanine synthase activity and cysteine synthase activity, is included within the meaning of "cysteine synthase".

The term "herbicide", as used herein, refers to a compound that may be used to kill or suppress the growth of at least one plant, plant cell, plant tissue or seed.

The term "inhibitor", as used herein, refers to a chemical substance that inactivates the enzymatic activity of cysteine synthase. The inhibitor may function by interacting directly with the enzyme, a cofactor of the enzyme, the substrate of the enzyme, or any combination thereof.

A polynucleotide may be "introduced" into a plant cell by any means, including transfection, transformation or transduction, electroporation, particle bombardment, agroinfection and the like. The introduced polynucleotide may be maintained in the cell stably if it is incorporated into a non-chromosomal autonomous replicon or integrated into the plant chromosome. Alternatively, the introduced polynucleotide may be present on an extra-chromosomal non-replicating vector and be transiently expressed or transiently active.

The "percent (%) sequence identity" between two polynucleotide or two polypeptide sequences is determined according to either the BLAST program (Basic Local Alignment Search Tool; Altschul and Gish (1996) *Meth Enzymol* 266:460-480 and Altschul (1990) *J Mol Biol* 215:403-410) in the Wisconsin Genetics Software Package (Devererreux *et al.* (1984) *Nucl Acid Res* 12:387), Genetics Computer Group (GCG), Madison, Wisconsin. (NCBI, Version 2.0.11, default settings) or using Smith Waterman Alignment (Smith and Waterman (1981) *Adv Appl Math* 2:482) as incorporated into GeneMatcher Plus<sup>TM</sup> (Paracel, Inc., <http://www.paracel.com/html/genematcher.html>; using the default settings and the version current at the time of filing). It is understood that for the purposes of

determining sequence identity when comparing a DNA sequence to an RNA sequence, a thymine nucleotide is equivalent to a uracil nucleotide.

“Plant” refers to whole plants, plant organs and tissues (e.g., stems, roots, ovules, stamens, leaves, embryos, meristematic regions, callus tissue, gametophytes, sporophytes, pollen, microspores and the like) seeds, plant cells and the progeny thereof.

By “polypeptide” is meant a chain of at least four amino acids joined by peptide bonds. The chain may be linear, branched, circular or combinations thereof. The polypeptides may contain amino acid analogs and other modifications, including, but not limited to glycosylated or phosphorylated residues.

The term “specific binding” refers to an interaction between cysteine synthase and a molecule or compound, wherein the interaction is dependent upon the primary amino acid sequence or the conformation of cysteine synthase.

As used herein, “sulfide” refers to inorganic sulfide or  $S^{2-}$ . The sulfide useful in the methods of the invention may be supplied by chemicals such as, but not limited to, hydrogen sulfide, sodium sulfide, potassium sulfide, rubidium sulfide, cesium sulfide, ammonium sulfide, beryllium sulfide, magnesium sulfide, calcium sulfide, strontium sulfide, barium sulfide and the like.

## Embodiments of the Invention

The present inventors have discovered that inhibition of cysteine synthase gene expression strongly inhibits the development and germination of plant seeds. Thus, the inventors are the first to demonstrate that cysteine synthase is a target for herbicides.

Accordingly, the invention provides methods for identifying compounds that inhibit cysteine synthase gene expression or activity. Such methods include ligand binding assays, assays for enzyme activity and assays for cysteine synthase gene expression. Any compound that is a ligand for cysteine synthase, other than its substrates, O-acetyl-L-serine and sulfide, or cofactors may have herbicidal activity. For the purposes of the invention, “ligand” refers to a molecule that will bind to a site on a polypeptide. The compounds identified by the methods of the invention are useful for the modulation of plant growth and development.

Thus, in one embodiment, the invention provides a method for identifying a compound as a candidate for a herbicide, comprising:

- a) contacting a cysteine synthase with said compound; and
- b) detecting the presence and/or absence of binding between said compound and said a cysteine synthase;

wherein binding indicates that said compound is a candidate for a herbicide.

By "cysteine synthase" is meant any enzyme that catalyzes the formation of cysteine from O-acetyl-serine and sulfide. The cysteine synthase may have the amino acid sequence of a naturally occurring cysteine synthase found in a plant, animal or  
 5 microorganism, or may have an amino acid sequence derived from a naturally occurring sequence. Preferably the cysteine synthase is a plant cysteine synthase.

By "plant cysteine synthase" is meant an enzyme that can be found in at least one plant, and which catalyzes the formation of cysteine from O-acetyl-serine and sulfide. The cysteine synthase may be from any plant, including both monocots and  
 10 dicots. In one embodiment, the cysteine synthase is an *Arabidopsis* cysteine synthase. *Arabidopsis* species include, but are not limited to, *Arabidopsis arenosa*, *Arabidopsis bursifolia*, *Arabidopsis cebennensis*, *Arabidopsis croatica*, *Arabidopsis griffithiana*, *Arabidopsis halleri*, *Arabidopsis himalaica*, *Arabidopsis korshinskyi*, *Arabidopsis lyrata*, *Arabidopsis neglecta*, *Arabidopsis pumila*, *Arabidopsis suecica*, *Arabidopsis thaliana* and *Arabidopsis wallichii*. Preferably, the *Arabidopsis* cysteine synthase is  
 15 from *Arabidopsis thaliana*, more preferably from *Arabidopsis thaliana* strain Columbia.

The cDNAs and amino acid sequences for the *A. thaliana* cysteine synthases are reported at GenBank and Embase accession numbers: emb | X80376 (*Atoacly*;  
 20 SEQ ID NO:1); emb | X84097 (*Atcys3A*; SEQ ID NO:2); emb | X81697 (*AtcysACSI*; SEQ ID NO:3); gb | AB024282 and emb | AJ010505 (*AtcysC1* or *oas5*; having both cysteine synthase and beta-cyanoalanine synthase activities; SEQ ID NOs: 4 and 5); gb | AB024284 (*AtcysD1* or *oas3*; SEQ ID NO:6); gb | X81973 (*mt ACSI*); gb | AB003041 (*cs26*); gb | AB02423 (*AtcysD2*); gb | X81698 (*cp ACS I*);  
 25 gb | AJ011044 (*oas6*); gb | AJ011976, gb | AJ011603, gb | AJ010505 (*oas5*), gb | BE039257 and gb | BE037810.

In addition, the sequences of cysteine synthase cDNAs and genes from a variety of other plants and organisms are publicly available. See, for example, gb | BE611445, gb | BE610004, gb | BE608380, gb | BE555595, gb | BE348173, gb | BE348127, gb | BE330806, gb | BE329795 and gb | BE020856 (*Glycine max* cysteine synthase cDNA); gb | AI563124 (watermelon cysteine synthase cDNA); gb | AF195239 (*Pyrus pyrifolia* cysteine synthase cDNA); gb | BE036966, gb | BE035521, gb | BE033421, gb | AI822724 and gb | AI822708 (*Mesembryanthemum crystallinum* cysteine synthase cDNA); gb | AI728743, gb | AI725847 and gb | AI725745 (*Gossypium hirsutum* cysteine synthase cDNA); gb | AW063019 (Sugar beet cysteine synthase cDNA); gb | AW164079 (*Lotus japonicus* cysteine synthase cDNA); gb | BE431668, gb | AW224722, gb | AW224770, gb | AW032102 and gb | AW031426 (*Lycopersicon esculentum* cysteine synthase); gb | D37963 and gb | D14722 (*Spinacia oleracea* cysteine synthase cDNA); gb | AI352787 (*Brassica napus* cysteine synthase cDNA); gb | AF044173 and gb | AF044172 (*Solanum tuberosum* cysteine synthase mRNA); gb | X85803 (*Zea mays* cysteine synthase cDNA); gb | AB040503 (*Allium tuberosum* cysteine synthase cDNA); gb | AF073695, gb | AF073696 and gb | AF073697 (*Oryza sativa* cysteine synthase *rsc1*, *rsc2* and *rsc3* cDNAs); gb | AW145239 (*Physcomitrella patens* cysteine synthase cDNA); gb | U19395 (*Emmericella nidulans* cysteine synthase (*cysB*) gene); gb | Z95395 (*Streptococcus gordonii* cysteine synthase gene); gb | X59595 (*Salmonella typhimurium* cysteine synthase *cys M* gene); gb | AW871292 and AW871235 (*Meloidogyne incognita* cysteine synthase mRNA); gb | AB028631 (*Entamoeba dispar* cysteine synthase 1 mRNA); gb | AB028632 (*Entamoeba dispar* cysteine synthase 2 mRNA); gb | AB006900, gb | AB000266 and gb | AB000265 (*Entamoeba histolytica* cysteine synthase mRNA); gb | AE000329 (*Escherichia coli* cysteine synthase gene); gb | AF246333 (*Acidithiobacillus ferrooxidans* cysteine synthase gene); gb | U93874 (*Bacillus subtilis* cysteine synthase); gb | AB028629 (*Clostridium perfringens* cysteine synthase); gb | AJ389044, gb | AJ389018 and gb | AJ388898 (*Medicago truncatula* cysteine synthase cDNA); gb | AF186381 (*Bacillus thermoleovorans* cysteine synthase cDNA); and gb | AW720774 (*Chlamydomonas reinhardtii* cysteine synthase cDNA). All of the above cysteine

synthase cDNA sequences may be used as probes to isolate cysteine synthase cDNAs or genes from additional organisms, and to synthesize cysteine synthase polypeptides.

In various embodiments, the cysteine synthase is from barnyard grass (*Echinochloa crus-galli*), crabgrass (*Digitaria sanguinalis*), green foxtail (*Setaria viridis*), perennial ryegrass (*Lolium perenne*), hairy beggarticks (*Bidens pilosa*), nightshade (*Solanum nigrum*), smartweed (*Polygonum lapathifolium*), velvetleaf (*Abutilon theophrasti*), common lambsquarters (*Chenopodium album* L.), *Brachiaria plantaginea*, *Cassia occidentalis*, *Ipomoea aristolochiaefolia*, *Ipomoea purpurea*, *Euphorbia heterophylla*, *Setaria* spp, *Amaranthus retroflexus*, *Sida spinosa*, *Xanthium strumarium* and the like.

Fragments of a plant cysteine synthase may be used in the methods of the invention. The fragments comprise at least 10 consecutive amino acids of a plant cysteine synthase. Preferably, the fragment comprises at least 15, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90 or at least 100 consecutive amino acids residues of a plant cysteine synthase. Most preferably, the fragment comprises at least 10 consecutive amino acid residues of an *Arabidopsis* cysteine synthase. Preferably, the fragment contains an amino acid sequence conserved among plant cysteine synthases. Such conserved fragments are identified in Rolland *et al.* (1993) *Biochem J* 293:829-833. Those skilled in the art could identify additional conserved fragments using sequence comparison software.

Polypeptides having at least 80% sequence identity with a plant cysteine synthase are also useful in the methods of the invention. Preferably, the sequence identity is at least 85%, more preferably the identity is at least 90%, most preferably the sequence identity is at least 95%.

In addition, it is preferred that the polypeptide has at least 50% of the activity of a plant cysteine synthase. More preferably, the polypeptide has at least 60%, at least 70%, at least 80% or at least 90% of the activity of a plant cysteine synthase. Most preferably, the polypeptide has at least 50%, at least 60%, at least 70%, at least 80% or at least 90% of the activity of the *A. thaliana* 'Columbia' cysteine synthase of SEQ ID NO:6.

Preferably, the activity of the polypeptide is compared to the activity of the *Arabidopsis* cysteine synthase polypeptide. For the purposes of the invention, cysteine synthase activity refers to the ability to convert O-acetyl-L-serine and sulfide



to L-cysteine and acetate. Methods for measuring cysteine synthase activity are known in the art and are described herein.

Thus, in another embodiment, the invention provides a method for identifying a compound as a candidate for a herbicide, comprising:

- 5           a)       contacting said compound with at least one polypeptide selected from the group consisting of: a plant cysteine synthase, a polypeptide comprising at least ten consecutive amino acids of a plant cysteine synthase, a polypeptide having at least 85% sequence identity with a plant cysteine synthase, and a polypeptide having at least 80% sequence identity with a plant cysteine synthase and at least 50% of the  
10       activity thereof; and
- b)       detecting the presence and/or absence of binding between said compound and said polypeptide;  
              wherein binding indicates that said compound is a candidate for a herbicide.

15           Any technique for detecting the binding of a ligand to its target may be used in the methods of the invention. Preferably, the ligand and target are combined in a buffer. Polypeptides and proteins that can reduce non-specific binding, such as BSA or protein extracts from cells that do not produce the target, may be included in binding assay.

20           Many methods for detecting the binding of a ligand to its target are known in the art, and include, but are not limited to the detection of an immobilized ligand-target complex or the detection of a change in the properties of a target when it is bound to a ligand. For example, in one embodiment, an array of immobilized candidate ligands is provided. The immobilized ligands are contacted with a cysteine  
25       synthase protein or a fragment or variant thereof, the unbound protein is removed and the bound cysteine synthase is detected. In a preferred embodiment, bound cysteine synthase is detected using a labeled binding partner, such as a labeled antibody. In a variation of this assay, cysteine synthase is labeled prior to contacting the  
30       immobilized candidate ligands. Preferred labels include fluorescent or radioactive moieties. Preferred detection methods include fluorescence correlation spectroscopy (FCS) and FCS-related confocal nanofluorimetric methods. See  
<http://www.evotec.de/technology>.

Once a compound is identified as a candidate for a herbicide, it can be tested for the ability to inhibit cysteine synthase enzyme activity. The compounds can be tested using either *in vitro* or cell based enzyme assays. Alternatively, a compound can be tested by applying it directly to a plant or plant cell, or expressing it therein, and monitoring the plant or plant cell for changes or decreases in growth, development, viability or alterations in gene expression.

Thus, in one embodiment, the invention provides a method for determining whether a compound identified as a herbicide candidate by an above method has herbicidal activity, comprising: contacting a plant or plant cells with said herbicide candidate and detecting the presence or absence of a decrease in the growth or viability of said plant or plant cells.

By decrease in growth, is meant that the herbicide candidate causes at least a 20% decrease in the growth of the plant or plant cells, as compared to the growth of the plants or plant cells in the absence of the herbicide candidate. By a decrease in viability is meant that at least 20% of the plants cells, or portion of the plant contacted with the herbicide candidate are nonviable. Preferably, the growth or viability will be at decreased by at least 40%. More preferably, the growth or viability will be decreased by at least 50%, 75% or at least 90% or more. Methods for measuring plant growth and cell viability are known to those skilled in the art. It is possible that a candidate compound may have herbicidal activity only for certain plants or certain plant species.

The ability of a compound to inhibit cysteine synthase activity can be detected using *in vitro* enzymatic assays in which the disappearance of a substrate or the appearance of a product is directly or indirectly detected. Cysteine synthase converts O-acetyl-L-serine and sulfide to cysteine and acetate. Methods for detection of these substrates and products, such as spectrophotometry, mass spectroscopy, thin layer chromatography (TLC), reverse phase HPLC and sulfide ion selective electrode assays are known to those skilled in the art. For example, cysteine can be measured by complexing with red ninhydrin (Gaitonde (1967) *Biochem J* 104:627-633), derivatization of the thiol group and identification by HPLC (Fahey *et al.* (1987) *Meth Enzymol* 143:85-96; Cooper *et al.* (1987) *Meth Enzymol* 143:141-143), determination of pyruvic acid after coupling with cysteine desulphydrase (Wedding (1987) *Meth Enzymol* 143:29-31) and by protein binding assays (Smith *et al.* (1987)

*Meth Enzymol* 143:144-148). In a preferred embodiment, cysteine synthase activity is measured using a ninhydrin assay in which cysteine concentration is measured by quantitating red ninhydrin complex at 550-560 nm. See, Schmidt (1990) *Methods in Plant Biochemistry* Vol. 3, Chapter 21, Dey *et al.*, Eds, Academic Press, London.

5 In the sulfide electrode assay, sulfide and calomel electrodes are attached to a signal buffer that serves as an impedance match. The sulfide concentration can be calculated from the potential measured by the electrode. See Hara *et al.* (1990) *Protein Expr Purif* 1:70-76.

Thus, the invention provides a method for identifying a compound as a  
10 candidate for a herbicide, comprising:

- a) contacting O-acetyl-L-serine and sulfide with cysteine synthase;
- b) contacting O-acetyl-L-serine and sulfide with cysteine synthase and said compound; and
- c) determining the concentration of cysteine, acetate, O-acetyl-L-serine and/or sulfide after the contacting of steps (a) and (b).

15

If a candidate compound inhibits cysteine synthase activity, a higher concentration of the substrates (O-acetyl-L-serine and sulfide) and a lower level of the products (cysteine and acetate) will be detected in the presence of the candidate compound (step b) than in the absence of the compound (step a).

20 Preferably the cysteine synthase is a plant cysteine synthase. Enzymatically active fragments of a plant cysteine synthase are also useful in the methods of the invention. For example, a polypeptide comprising at least 100 consecutive amino acid residues of a plant cysteine synthase may be used in the methods of the invention. In addition, a polypeptide having at least 80%, 85%, 90%, 95%, 98% or at  
25 least 99% sequence identity with a plant cysteine synthase may be used in the methods of the invention. Also, polypeptides having at least 80% sequence identity with at least 15 consecutive amino acid residues of a plant cysteine synthase are also useful in the methods of the invention. Preferably, the polypeptide has at least 80% sequence identity with a plant cysteine synthase and at least 50%, 75%, 90% or at  
30 least 95% of the activity thereof.

Thus, the invention provides a method for identifying a compound as a candidate for a herbicide, comprising:

- a) contacting O-acetyl-L-serine and sulfide with a polypeptide selected from the group consisting of: a polypeptide having at least 85%  
5 sequence identity with a plant cysteine synthase, a polypeptide having at least 80% sequence identity with a plant cysteine synthase and at least 50% of the activity thereof, and a polypeptide comprising at least 100 consecutive amino acids of a plant cysteine synthase;
- b) contacting O-acetyl-L-serine and sulfide with said polypeptide  
10 and said compound; and
- c) determining the concentration of cysteine, acetate, O-acetyl-L-serine and/or sulfide after the contacting of steps (a) and (b).

Again, if a candidate compound inhibits cysteine synthase activity, a higher concentration of the substrate (O-acetyl-L-serine and sulfide) and a lower level of the product (cysteine and acetate) will be detected in the presence of the candidate  
15 compound (step b) than in the absence of the compound (step a).

For the *in vitro* enzymatic assays, cysteine synthase protein and derivatives thereof may be purified from a plant or may be recombinantly produced in and purified from a plant, bacteria, or eukaryotic cell culture. Preferably these proteins are produced using a baculovirus or *E. coli* expression system. See, for example,  
20 Saito *et al.* (1992) *Proc Natl Acad Sci* 89:8078-8082. Methods for the purification of cysteine synthase from potato tubers, bell pepper, spinach and *Salmonella typhimurium* are described in Ishizawa *et al.* (1991) *Plant and Cell Physiol* 41:200-208; Romer *et al.* (1992) *J Biol Chem* 267:17966-17970; Yamaguchi *et al.* (1998) *Bioscience, Biotechnology and Biochemistry* 62:501-507; and Hara *et al.* (1990)  
25 *Protein Expr Purif* 1:70-76, respectively. Other methods for the purification of cysteine synthase proteins and polypeptides are known to those skilled in the art.

As an alternative to *in vitro* assays, the invention also provides plant and plant cell based assays. In one embodiment, the invention provides a method for identifying a compound as a candidate for a herbicide, comprising:

- a) measuring the expression of cysteine synthase in a plant or plant cell in  
30 the absence of said compound;

- b) contacting a plant or plant cell with said compound and measuring the expression of cysteine synthase in said plant or plant cell;
- c) comparing the expression of cysteine synthase in steps (a) and (b).

5 A reduction in cysteine synthase expression indicates that the compound is a herbicide candidate. In one embodiment, the plant or plant cell is an *Arabidopsis thaliana* plant or plant cell.

Expression of cysteine synthase can be measured by detecting cysteine synthase primary transcript or mRNA, cysteine synthase polypeptide or cysteine  
10 synthase enzymatic activity. Methods for detecting the expression of RNA and proteins are known to those skilled in the art. See, for example, *Current Protocols in Molecular Biology* Ausubel *et al.*, eds., Greene Publishing and Wiley-Interscience, New York, 1995. The method of detection is not critical to the invention. Methods for detecting cysteine synthase RNA include, but are not limited to amplification  
15 assays such as quantitative PCR, and/or hybridization assays such as Northern analysis, dot blots, slot blots, in-situ hybridization, transcriptional fusions using a cysteine synthase promoter fused to a reporter gene, bDNA assays and microarray assays.

Methods for detecting protein expression include, but are not limited to,  
20 immunodetection methods such as Western blots, His Tag and ELISA assays, polyacrylamide gel electrophoresis, mass spectroscopy and enzymatic assays. Also, any reporter gene system may be used to detect cysteine synthase protein expression. For detection using gene reporter systems, a polynucleotide encoding a reporter protein is fused in frame with cysteine synthase, so as to produce a chimeric  
25 polypeptide. Methods for using reporter systems are known to those skilled in the art. Examples of reporter genes include, but are not limited to, chloramphenicol acetyltransferase (Gorman *et al.* (1982) *Mol Cell Biol* 2:1104; Prost *et al.* (1986) *Gene* 45:107-111),  $\beta$ -galactosidase (Nolan *et al.* (1988) *Proc Natl Acad Sci USA* 85:2603-2607), alkaline phosphatase (Berger *et al.* (1988) *Gene* 66:10), luciferase (De Wet *et al.* (1987) *Mol Cell Biol* 7:725-737),  $\beta$ -glucuronidase (GUS), fluorescent proteins,  
30 chromogenic proteins and the like. Methods for detecting cysteine synthase activity are described above.

Chemicals, compounds or compositions identified by the above methods as modulators of cysteine synthase expression or activity can then be used to control plant growth. For example, compounds that inhibit plant growth can be applied to a plant or expressed in a plant, in order to prevent plant growth. Thus, the invention provides a method for inhibiting plant growth, comprising contacting a plant with a compound identified by the methods of the invention as having herbicidal activity.

Herbicides and herbicide candidates identified by the methods of the invention can be used to control the growth of undesired plants, including both monocots and dicots. Examples of undesired plants include, but are not limited to barnyard grass (*Echinochloa crus-galli*), crabgrass (*Digitaria sanguinalis*), green foxtail (*Setaria viridis*), perennial ryegrass (*Lolium perenne*), hairy beggarticks (*Bidens pilosa*), nightshade (*Solanum nigrum*), smartweed (*Polygonum lapathifolium*), velvetleaf (*Abutilon theophrasti*), common lambsquarters (*Chenopodium album* L.), *Brachiaria plantaginea*, *Cassia occidentalis*, *Ipomoea aristolochiaefolia*, *Ipomoea purpurea*, *Euphorbia heterophylla*, *Setaria spp*, *Amaranthus retroflexus*, *Sida spinosa*, *Xanthium strumarium* and the like.

## EXPERIMENTAL

### Plant Growth Conditions

Unless, otherwise indicated, all plants were grown Scotts Metro-Mix™ soil (the Scotts Company) or a similar soil mixture in an environmental growth room at 22°C, 65% humidity, 65% humidity and a light intensity of  $\sim 100 \mu\text{-E m}^{-2} \text{ s}^{-1}$  supplied over 16 hour day period.

### Seed Sterilization

All seeds were surface sterilized before sowing onto phytagel plates using the following protocol.

1. Place approximately 20-30 seeds into a labeled 1.5 ml conical screw cap tube. Perform all remaining steps in a sterile hood using sterile technique.
2. Fill each tube with 1ml 70% ethanol and place on rotisserie for 5 minutes.

3. Carefully remove ethanol from each tube using a sterile plastic dropper; avoid removing any seeds.
4. Fill each tube with 1ml of 30% Clorox and 0.5% SDS solution and place on rotisserie for 10 minutes.
5. Carefully remove bleach/SDS solution.
6. Fill each tube with 1ml sterile dI H<sub>2</sub>O; seeds should be stirred up by pipetting of water into tube. Carefully remove water. Repeat 3 to 5 times to ensure removal of Clorox/SDS solution.
7. Fill each tube with enough sterile dI H<sub>2</sub>O for seed plating (~200-400 µl). Cap tube until ready to begin seed plating.

#### *Plate Growth Assays*

Surface sterilized seeds were sown onto plate containing 40 ml half strength sterile MS medium (no sucrose) and 1% Phytigel using the following protocol:

1. Using pipette man and 200 µl tip, carefully fill tip with seeds and 0.1% agarose solution. Place 10 seeds across the top of the plate, about ¼ in down from the top edge of the plate.
2. Place plate lid ¾ of the way over the plate and allow to dry for 30 minutes or until agarose solution is dry. It is important to allow agarose solution to dry completely before sealing up plates in order to prevent contamination.
3. Using sterile micropore tape, seal the edge of the plate where the top and bottom meet.
4. Place plates stored in a vertical rack in the dark at 4°C for three days.
5. Three days after sowing, the plates transferred into a growth chamber with a day and night temperature of 22 and 20 °C, respectively, 65% humidity and a light intensity of ~100 µ-E m<sup>-2</sup> s<sup>-1</sup> supplied over 16 hour day period.
6. Beginning on day 3, daily measurements are carried out to track the seedlings development until day 14. Seedlings are harvested on day 14 (or when root length reaches 6 cm) for root and rosette analysis.

## Example 1

## Construction of a Transgenic Plant expressing the Driver

The "Driver" is an artificial transcription factor comprising a chimera of the DNA-binding domain of the yeast GAL4 protein (amino acid residues 147) fused to two tandem activation domains of herpes simplex virus protein VP16 (amino acid residues 413-490). Schwechheimer *et al.* (1998) *Plant Mol Biol* 36:195-204. This chimeric driver is a transcriptional activator specific for promoters having GAL4 binding sites. Expression of the driver is controlled by two tandem copies of the constitutive CaMV 35S promoter.

The driver expression cassette was introduced into *Arabidopsis thaliana* by agroinfection. Transgenic plants that stably expressed the driver transcription factor were obtained.

## Example 2

## Construction of Cysteine Synthase Antisense Expression Cassettes in a Binary Vector

Portions of two *Arabidopsis thaliana* cysteine synthase cDNAs were ligated into the PacI /AscI sites of the *E.coli/Agrobacterium* binary vector PGT3.2 in the antisense orientation. This placed transcription of the cysteine synthase antisense RNAs under the control of an artificial promoter that is active only in the presence of the driver transcription factor described above. The artificial promoter contains four contiguous binding sites for the GAL4 transcriptional activator upstream of a minimal promoter comprising a TATA box.

The ligated DNA was transformed into *E.coli*. Kanamycin resistant clones were selected and purified. DNA was isolated from each clone and characterized by PCR and sequence analysis. pPG237 expresses an *A. thaliana* cysteine synthase antisense RNA that is complementary to 761/772 nucleotides of the *AtcysC1* mRNA (nucleotides 1348 to 578 of gb | AB024282) and to 760/772 of the *Arabidopsis thaliana* cysteine synthase reported as emb | AJ010505. pPG244 expresses an *A. thaliana* cysteine synthase antisense RNA that is complementary to 554/556 nucleotides of the *Atoacyl* mRNA (nucleotides 1226 to 671 of emb | X80376), 552/556 nucleotides of the *Atcys3A* mRNA (nucleotides 1263 to 708 of



emb | X84097) and 532/562 nucleotides of the *AtcytACS1* mRNA (nucleotides 1259 to 708 of emb | X81697).

The antisense expression cassette and a constitutive barnase expression cassette are located between right and left T-DNA borders. Thus, the antisense expression cassettes can be transferred into a recipient plant cell by agroinfection.

### Example 3

#### Transformation of *Agrobacterium* with the Target Expression Cassette

pPG237 and pPG244 were transformed into *Agrobacterium tumefaciens* by electroporation. Transformed *Agrobacterium* colonies were isolated using Basta selection. DNA was prepared from purified Basta resistant colonies and the inserts were amplified by PCR and sequenced to confirm sequence and orientation. The clones were stored as a frozen glycerol stock.

### Example 4

#### Construction of an *Arabidopsis* cysteine synthase Antisense Target Plants

The cysteine synthase target expression cassettes were introduced into *Arabidopsis thaliana* wild-type plants by the following method. Five days prior to agroinfection, the primary inflorescence of *Arabidopsis thaliana* plants grown in 2.5 inch pots were clipped in order enhance the emergence of secondary bolts.

At two days prior to agroinfection, 5 ml LB broth (10 g/L Peptone, 5 g/L Yeast extract, 5 g/L NaCl, pH 7.0 plus 25 mg/L kanamycin added prior to use) was inoculated with a clonal glycerol stock of *Agrobacterium* carrying pPG237 or pPG244. The cultures were incubated overnight at 28°C at 250 rpm until the cells reached stationary phase. The following morning, 200 ml LB in a 500 ml flask was inoculated with 500 µl of the overnight culture and the cells were grown to stationary phase by overnight incubation at 28°C at 250 rpm. The cells were pelleted by centrifugation at 8000 rpm for 5 minutes. The supernatant was removed and excess media was removed by setting the centrifuge bottles upside down on a paper towel for several minutes. The cells were then resuspended in 500 ml infiltration medium (autoclaved 5% sucrose) and 250 µl/L Silwet L-77™ (84% polyalkyleneoxide

modified heptamethyltrisiloxane and 16% allyloxypolyethyleneglycol methyl ether), and transferred to a one liter beaker.

The previously clipped *Arabidopsis* plants were dipped into the *Agrobacterium* suspension so that all above ground parts were immersed and agitated gently for 10 seconds. The dipped plants were then cover with a tall clear plastic dome in order to maintain the humidity, and returned to the growth room. The following day, the dome was removed and the plants were grown under normal light conditions until mature seeds were produced. Mature seeds were collected and stored desiccated at 4 °C.

Transgenic *Arabidopsis* T1 seedlings were selected using glufosinate treatment. Approximately 70 mg seeds from an agrotransformed plant were mixed approximately 4:1 with sand and placed in a 2 ml screw cap cryo vial.

The surface of the seeds was sterilized using the chlorine gas method. Briefly, the open vials were placed in a vacuum desiccator in a safety hood. A glass beaker containing 200 ml 5.25% sodium hypochlorite solution was placed in the desiccator. Two ml concentrated HCl was added to the hypochlorite solution and the cover was placed on the desiccator. Vacuum was applied briefly to seal the dessicator, and the seeds were left in the desiccator overnight.

One vial of sterilized seeds was then sown in a cell of an 8 cell flat. The flat was covered with a dome, stored at 4°C for 3 days, and then transferred to a growth room. The domes were removed when the seedlings first emerged. After the emergence of the first primary leaves, the flat was sprayed uniformly with a 1:3000 dilution of Liberty™ (AgrEvo; 11.3% glufosinate) in water, 0.005% Silwet (50 µl/L) until the leaves were completely wetted. The spraying was repeated for the following two days.

Ten days after the first spraying resistant plants were transplanted to 2.5 inch round pots containing moistened sterile potting soil. The transplants were then sprayed with herbicide and returned to the growth room. These herbicide resistant plants represent stably transformed T1 plants. Mature T1 plants are then dried and harvested for T2 seeds.

Example 5  
Effect of cysteine synthase Antisense Expression  
in *Arabidopsis* Seedlings

5        The cysteine synthase target plants from the transformed plant lines obtained in Example 4 were crossed with the *Arabidopsis* transgenic driver line described above. The resulting F1 seeds were then subjected to a PGI plate assay to observe seedling growth over a 2-week period. Seedlings were inspected daily for growth and development. During this period, approximately half of seeds derived from one of the

10       pPG237 cysteine synthase antisense target lines developed abnormal cotyledons and showed reduced root growth. This pPG237 line also showed a 76% reduction in total leaf area and a 73% reduction in root length as compared to wild-type plants. One of the pPG244 cysteine synthase antisense target lines was white and failed to germinate, apparently representing an embryo lethal phenotype, while about half of the plants of

15       a second line exhibited mild chlorosis and developmental abnormalities. The results are summarized in Table 1.

**Table 1**  
**Phenotypes of plants expressing cysteine synthase antisense RNA**

20

Construct	No. Wild Type	No. Abnormal	$\chi^2$ Value <sup>a</sup>	Probability <sup>a</sup>
pPG237	7	3	0.900	0.348
pPG244	4	6	0.400	0.527

<sup>a</sup>Chi-square and P values (0.05) were obtained to evaluate the hypothesis that chlorosis and wild-type phenotypes are segregating in a 1:1 ratio.

25       The clear 1:1 segregation ration observed demonstrates that the antisense expression of these genes encoding cysteine synthases results in the inhibition of germination and causes abnormal development. Thus, cysteine synthases are essential genes for normal plant growth and development. The fact that progeny derived from other lines selected from the pPG237 and pPG244 transformations did not exhibit any

abnormal phenotypes is not unexpected since it well known that antisense expression does not work equally well in all independently transformed lines containing the same construct.

5

### Example 6

#### Assay for Inhibitors of Cysteine Synthase Activity

The enzymatic activity of the cysteine synthase of SEQ ID NO:6 is determined in the presence and absence of candidate inhibitors in a 0.5 ml reaction mixture containing 50 mM potassium phosphate, pH 8.0, 4 mM sodium sulfide, 12.5 mM O-acetyl-L-serine and 10 µg cysteine synthase. The reaction mixture is incubated at 30°C for thirty minutes and terminated by the addition of 0.1 ml 7.5% TCA. The amount of cysteine in the resulting mixture is determined spectrophotometrically at 560 nm by the acid-ninhydrin method of Gaitonde (1967) *Biochem J* 104:627-633.

15

While the foregoing describes certain embodiments of the invention, it will be understood by those skilled in the art that variations and modifications may be made and still fall within the scope of the invention.

## CLAIMS

1. A method for identifying a compound as a candidate for a herbicide, comprising:
  - a) contacting a cysteine synthase with said compound; and
  - b) detecting the presence and/or absence of binding between said compound and said a cysteine synthase;wherein binding indicates that said compound is a candidate for a herbicide.
2. The method of claim 1, wherein said cysteine synthase is a plant cysteine synthase.
3. The method of claim 2, wherein said cysteine synthase is an *Arabidopsis* cysteine synthase.
4. The method of claim 3, wherein said *Arabidopsis* cysteine synthase is a protein selected from the group consisting of: SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5 and SEQ ID NO:6.
5. A method for determining whether a compound identified as a herbicide candidate by the method of claim 1 has herbicidal activity, comprising: contacting a plant or plant cells with said herbicide candidate and detecting the presence or absence of a decrease in growth or viability of said plant or plant cells.
6. A method for identifying a compound as a candidate for a herbicide, comprising:
  - a) contacting said compound with at least one polypeptide selected from the group consisting of: an amino acid sequence comprising at least ten consecutive amino acids of a plant cysteine synthase, an amino acid sequence having at least 85% sequence identity with a plant cysteine synthase, and an amino acid sequence having at least 80% sequence identity with a plant cysteine synthase and at least 50% of the activity thereof; and

b) detecting the presence and/or absence of binding between said compound and said polypeptide;

wherein binding indicates that said compound is a candidate for a herbicide.

7. A method for determining whether a compound identified as a herbicide candidate by the method of claim 6 has herbicidal activity, comprising: contacting a plant or plant cells with said herbicide candidate and detecting the presence or absence of a decrease in growth or viability of said plant or plant cells.

8. A method for identifying a compound as a candidate for a herbicide, comprising:

a) contacting O-acetyl-L-serine and sulfide with a cysteine synthase in the absence of said compound;

b) contacting O-acetyl-L-serine and sulfide with a cysteine synthase in the presence of said compound;

c) determining the concentration of O-acetyl-L-serine, sulfide, cysteine and/or acetate after the contacting of steps (a) and (b).

9. The method of claim 8, wherein said cysteine synthase is a plant cysteine synthase.

10. The method of claim 9, wherein said cysteine synthase is an *Arabidopsis* cysteine synthase.

11. The method of claim 10, wherein said *Arabidopsis* cysteine synthase is a protein selected from the group consisting of: SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5 and SEQ ID NO:6.

12. A method for determining whether a compound identified as a herbicide candidate by the method of claim 8 has herbicidal activity, comprising: contacting a plant or plant cells with said herbicide candidate and detecting the presence or absence of a decrease in growth or viability of said plant or plant cells.

13. A method for identifying a compound as a candidate for a herbicide, comprising:

- a) contacting O-acetyl-L-serine and sulfide with a polypeptide selected from the group consisting of: an amino acid sequence comprising at least 100 consecutive amino acids of a plant cysteine synthase, an amino acid sequence having at least 85% sequence identity with a plant cysteine synthase, and an amino acid sequence having at least 80% sequence identity with a plant cysteine synthase and at least 50% of the activity thereof, in the absence of said compound;
- b) contacting O-acetyl-L-serine and sulfide with said polypeptide in the presence of said compound;
- c) determining the concentration of O-acetyl-L-serine, sulfide, cysteine and/or acetate remaining after the contacting of steps (a) and (b).

14. A method for determining whether a compound identified as a herbicide candidate by the method of claim 13 has herbicidal activity, comprising: contacting a plant or plant cells with said herbicide candidate and detecting the presence or absence of a decrease in growth or viability of said plant or plant cells.

15. A method for identifying a compound as a candidate for a herbicide, comprising:

- a) measuring the expression of cysteine synthase in a plant or plant cell in the absence of said compound;
- b) contacting a plant or plant cell with said compound and measuring the expression of cysteine synthase in said plant or plant cell;
- 5 c) comparing the expression of cysteine synthase in steps (a) and (b).

16. The method of claim 15 wherein said plant or plant cell is an *Arabidopsis* plant or plant cell.

17. The method of claim 15, wherein the expression of cysteine synthase is measured by detecting cysteine synthase mRNA.

18. The method of claim 15, wherein the expression of cysteine synthase is measured by detecting cysteine synthase polypeptide.

19. A method for determining whether a compound identified as a herbicide candidate by the method of claim 15 has herbicidal activity, comprising: contacting a plant or plant cells with said herbicide candidate and detecting the presence or absence of a decrease in growth or viability of said plant or plant cells.

20. The method of claim 19 wherein said plant or plant cell is an *Arabidopsis* plant or plant cell.



## SEQUENCE LISTING

<110> Kloti, Andreas  
 Woessner, Jeffrey  
 Zayed, Adel  
 Gorlach, Jorn  
 Boyes, Doug  
 Davis, Keith  
 Hamilton, Carol  
 Ascenzi, Robert  
 Hoffman, Neil

<120> METHODS FOR THE IDENTIFICATION OF  
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&lt;213&gt; Arabidopsis thaliana

&lt;400&gt; 5

Met	Ala	Ser	Val	Ser	Arg	Arg	Leu	Leu	Arg	Arg	Glu	Thr	Ile	Pro	Cys		
1				5					10					15			
Phe	Ser	His	Thr	Val	Arg	Lys	Leu	Phe	Ser	Thr	Val	Gly	Ser	Pro	Ser		
		20						25				30					
Phe	Ala	Gln	Arg	Leu	Arg	Asp	Leu	Pro	Lys	Asp	Phe	Pro	Ser	Thr	Asn		
	35						40					45					
Ala	Lys	Arg	Asp	Ala	Ser	Leu	Leu	Ile	Gly	Lys	Thr	Pro	Leu	Val	Phe		

50		55		60	
Leu Asn Lys Val Thr Glu Gly Cys Glu Ala Tyr Val Ala Ala Lys Gln					
65		70		75	80
Glu His Phe Gln Pro Thr Cys Ser Ile Lys Asp Arg Pro Ala Ile Ala					
	85		90		95
Met Ile Ala Asp Ala Glu Lys Lys Lys Leu Ile Ile Pro Gly Lys Thr					
	100		105		110
Thr Leu Ile Glu Pro Thr Ser Gly Asn Met Gly Ile Ser Leu Ala Phe					
	115		120		125
Met Ala Ala Met Lys Gly Tyr Arg Ile Ile Met Thr Met Pro Ser Tyr					
	130		135		140
Thr Ser Leu Glu Arg Arg Val Thr Met Arg Ser Phe Gly Ala Glu Leu					
145		150		155	160
Val Leu Thr Asp Pro Ala Lys Gly Met Gly Gly Thr Val Lys Lys Ala					
	165		170		175
Tyr Asp Leu Leu Asp Ser Thr Pro Asp Ala Phe Met Cys Gln Gln Phe					
	180		185		190
Ala Asn Pro Ala Asn Thr Gln Ile His Phe Asp Thr Thr Gly Pro Glu					
	195		200		205
Ile Trp Glu Asp Thr Leu Gly Asn Val Asp Ile Phe Val Met Gly Ile					
	210		215		220
Gly Ser Gly Gly Thr Val Ser Gly Val Gly Arg Tyr Leu Lys Ser Lys					
225		230		235	240
Asn Pro Asn Val Lys Ile Tyr Gly Val Glu Pro Ala Glu Ser Asn Ile					
	245		250		255
Leu Asn Gly Gly Lys Pro Gly Pro His Ala Ile Thr Gly Asn Gly Val					
	260		265		270
Gly Phe Lys Pro Glu Ile Leu Asp Met Asp Val Met Glu Ser Val Leu					
	275		280		285
Glu Val Ser Ser Glu Asp Ala Ile Lys Met Ala Arg Glu Leu Ala Leu					
	290		295		300
Lys Glu Gly Leu Met Val Gly Ile Ser Ser Gly Ala Asn Thr Val Ala					
305		310		315	320
Ala Ile Arg Leu Ala Lys Met Pro Glu Asn Lys Gly Lys Leu Ile Val					
	325		330		335
Thr Ile His Ala Ser Phe Gly Glu Arg Tyr Leu Ser Ser Val Leu Phe					
	340		345		350
Asp Glu Leu Arg Lys Glu Ala Glu Glu Met Lys Pro Val Ser Val Asp					
	355		360		365

&lt;210&gt; 6

&lt;211&gt; 324

&lt;212&gt; PRT

&lt;213&gt; Arabidopsis thaliana

&lt;400&gt; 6

Met Glu Glu Asp Arg Cys Ser Ile Lys Asp Asp Ala Thr Gln Leu Ile		
1	5	10
Gly Asn Thr Pro Met Val Tyr Leu Asn Asn Ile Val Asp Gly Cys Val		
	20	25
Ala Arg Ile Ala Ala Lys Leu Glu Met Met Glu Pro Cys Ser Ser Val		
	35	40
Lys Glu Arg Ile Ala Tyr Gly Met Ile Lys Asp Ala Glu Asp Lys Gly		
	50	55
Leu Ile Thr Pro Gly Lys Ser Thr Leu Ile Glu Ala Thr Ser Gly Asn		
65	70	75
Thr Gly Ile Gly Leu Ala Phe Ile Gly Ala Ala Lys Gly Tyr Lys Val		
	85	90
Val Leu Thr Met Pro Ser Ser Met Ser Leu Glu Arg Lys Ile Ile Leu		

